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## Constitutively Active Receptors

File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS A GROUP I					
MSHR_mouse	melanocyte-stimulating hormone	TMII	92 VSIIVL <del>G</del> TILL K	adenylyl cyclase activity/ HEK293, stably transfected	(Robbins, Nadeau et al. 1993)
CLASS A GROUP II	MSH				
SHB_human	5-hydroxytryptamine <sub>1B</sub>	C-terminus of IC3	313 RERKATKTLGI K,R,Q	binding of [ <sup>35</sup> S]GTP[S] / CHO-KI	(Pauwels, Gouble 1999)
SH2A_human	5-hydroxytryptamine <sub>2A</sub>	C-terminus of IC3	322 NEQKACKVLGI K	IP production / COS-7	(Egan, Herrick-Davis et al. 1998)
2H2C_rat	5-hydroxytryptamine <sub>2C</sub>	C-terminus of IC3	312 NEDDASKVLGI L	PI hydrolysis / COS-7	(Herrick-Davis, Egan et al. 1997)

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CLASS A GROUP II				
<b>α1AB_human</b>	$\alpha_{1B}$ -adrenergic alpha 1B-AR	TMDI junction between TMDIII and IC2	63 <b>FAIVGNILVIL</b> A 142 <b>CAISIDRYIGV</b> A	IP / COS-7 (Scheer, Fanelli et al. 1997)
<b>α1AB_human</b>	$\alpha_{1B}$ -adrenergic alpha 1B-AR	TMDI junction between TMDIII and IC2	143 <b>CARISIDRYIGV</b> K 128 <b>AVDVLLCTASI</b> F	IP / COS-7 (Scheer, Costa et al. 2000)
<b>α1AB_human</b>	$\alpha_{1B}$ -adrenergic	carboxyl end of IC3 TMV	293 <b>REKKAA<del>K</del>KTTLGI</b> E 204 <b>EFPFYALFSSLG</b> V	IP / COS-1 (Perez, Hwa et al. 1999)
<b>α1AB_human</b>	$\alpha_{1B}$ -adrenergic	C-terminal IC3	293 <b>SREKKAA<del>K</del>KT</b> X=19 different substitutions K H L	IP / COS-7 (Hwa, Gaivin et al. 1997)
<b>α1AB_human</b>	$\alpha_{1B}$ -adrenergic	C-terminus IC3	288 293 <b>KFSREKKAA<del>K</del>KTTLGI</b> K H L	PI / COS-7 (Kjelsberg, Coletchchia et al. 1992)
<b>A2AA_human</b>	$\alpha_2$ C10-adrenergic alpha-2AAR	C-terminal IC3 loop	373 (348?) <b>EKRFTFVLAV</b> X=F,A,C,E;K 360 <b>SLVK<del>E</del>KKAAARTLS</b> A	PI hydrolysis / rat fibroblast adenylyl cyclase inhibition / HEK293 (Ren, Kurose et al. 1993)
<b>ACM1_human</b>	muscarinic M1	C-terminal IC3 loop junction	390 <b>KKVTRTIL<del>A</del></b> 1-4 A inserted	(Högger, Shockley et al. 1995)
<b>ACM2-human</b>	muscarinic acetylcholine M2	junction of IC3 and TMV	IP production, inhibition of cAMP production / COS-7 (Liu, Blin et al. 1996)	

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CLASS A GROUP II				
ACM3_rat	m3 muscarinic (rat)	TMVI	507 TWT <sup>S</sup> PYNNIMVLVNT	IP / COS-7 (Blümli, Mutschler et al. 1994)
ACM5_human	m5 muscarinic muscarinic acetylcholine M5	N-terminus to TMII	chimera composed of m2 1-69 m5 77-445 m2 391-466	β-gal / NIH 3T3 (Burstein, Spalding et al. 1996)
ACM5_human	m5 muscarinic muscarinic acetylcholine M5	TMVI	451 AILLA <sup>M</sup> FIITW <sup>L</sup> TPYNI <sup>H</sup> MVLV <sup>S</sup> T <sup>C</sup> T <sup>F</sup>	β-gal; radioligand binding / NIH-3T3 (Spalding, Burstein et al. 1998)
ACMS_human	m5 muscarinic muscarinic acetylcholine M5	junction of TMVI and EC3	465 YNIMVLV <sup>S</sup> TFCDKCV X=V,F,R,K,+more	β-gal; radioligand binding / NIH-3T3 (Spalding, Burstein et al. 1997)
B1AR_human	β <sub>1</sub> -adrenergic	C-terminus	389 RKAFQGLLCA <sup>R</sup>	adenylyl cyclase; agonist binding / CHW (Mason, Moore et al. 1999)
B2AR_human	β <sub>2</sub> -adrenergic beta-2AR	C-terminal IC3 loop	266 272 FCLKEH <sup>I</sup> KAL <sup>K</sup> TKLGI <sup>A</sup> SR K A	adenylyl cyclase activation; agonist binding affinity / COS-7 or CHO (Samama, Cotecchia et al. 1993); (Leffkowitz, Cotecchia et al. 1993)
DADR_human	dopamine D1A	carboxyl terminal IC3	264 SFKMS <sup>I</sup> EKKR <sup>K</sup> ETKV <sup>A</sup> LKT 288 from D1B receptor APDTSIKKETKV <sup>L</sup> KT	adenylyl cyclase; cAMP accumulation / HEK293 (Charpentier, Jarvie et al. 1996)
DADR_human	dopamine D1	TMVI	286 FVCCWL <sup>A</sup> PFFFIL	cAMP accumulation / COS-7 (Cho, Taylor et al. 1996)
HH2R_rat	histamine H <sub>2</sub>	IC2	115 FMISLD <sup>N</sup> RYCAV <sup>A</sup>	cAMP production / HEK-293 (Alewijse, Timmerman et al. 2000)

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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS A GROUP III					
OPSD_human	opsin	TMII	90 FVNLLGGFTSTLY	transducin;	(Rim and Oprian 1995)
	rhodopsin	TMIII	D 113 GCNLEGFFAT	phosphorylation by rhodopsin kinase / COS	
		TMVII	Q 292 296 MTIPIAFFAKSAAIY		
			E G, E, M		
			<sup>292</sup> Ala neutral a.a converted to carboxylate and competes with <sup>113</sup> Glu for salt bridge with <sup>296</sup> Lys		
OPSD_human	opsin	TMII	134 VVLAI <del>E</del> RYVVV I, Q, S	transducin; radioligand binding / COS	(Acharya and Karnik 1996)
	rhodopsin	TMIV	257 RMVIIIMVIAFL Y, N	transducin, GTP-S uptake / COS	(Han, Smith et al. 1998)
OPSD_human	opsin	TMVI	plus TM3 296 PAFFAKSAIY G	transducin; radioligand binding / COS	(Govardhan and Oprian 1994); (Cohen, Yang et al. 1993)
	rhodopsin		X=E,M natural mutants + 10 different a.a. substitutions		
			disrupts critical salt bridge between <sup>296</sup> Lys(TMVII) and <sup>113</sup> Glu(TMIII)		
		IC2	134 VVLAI <del>E</del> RYVVV Q		(Cohen, Yang et al. 1993)

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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS A GROUP IV					
BRB2_human	bradykinin B, B2 bradykinin BK-2	TMII TMI	113 A I I S M A Y S S I A 256 L L F I I C W L P P Q I F	IP production / COS-7	(Marie, Koch et al. 1999)

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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS A GROUP V					
AG2R_rat	AT <sub>1A</sub>	TMII	111 ASVSPNLVASY	phospholipase C, IP production / COS-7	(Groblewski; Magret et al. 1997)
	Type 1A angiotensin II		A disrupts Asn (TMII)- Tyr (TMVII) interaction.		
AG2R_rat	AT <sub>1A</sub>	C-terminus of TM7	305 LFYCFGLKKEK	IP production / HEK- 293; intracellular Ca <sup>2+</sup> mobilization / CHO	(Pannier, Bardin et al. 2000)
	Type 1A angiotensin II	other multiple mutations	Q 51 LVIVWAGFERNATITITISYKAVAA LWVWVTAFAEKRTINAFLNLAVA (K above conflicts with SWISS-PROT database)	PI production; phospholipase C stimulation / COS-7	(Amatudia, Draga Graonic et al. 1995)
FMLR_human	formy(methionyl)leucylphenylalanine (FMLPR)	IC1	138 ACISVDRYIAIVH V	IP production; Ca <sup>2+</sup> mobilization and actin polymerization / NIH 3T3	(Burger, Burger et al. 1999)
IL8B_human	interleukin-8 receptor B	IC2	564 MATNKDTKIAKK G	cAMP production / HEK293	(Kudo, Osuga et al. 1996)
CXCR-2 chemokine		IC3	578 ILLIFTDFTCMAG	cAMP production / COS-7	(Shenker, Lue et al. 1993)
LSHR_human	luteinizing hormone (LH)	TMVI	571-577 KIAKKMATTIILIFTDFCM I I I	cAMP production / COS-7	(Kosugi, Van Dop et al. 1995)
LSHR_human	luteinizing hormone (LH)	TM6	556 ILIFTDFTCMAG G, Y	cAMP production / HEK 293T	(Bradbury, Kawate et al. 1997; Bradbury and Menon 1999)
LSHR_rat	luteinizing hormone / human chorionic gonadotropin (LH/hCG)	TMVI	128 KVLSTIDYXNMP A, K, H	adenylyl cyclase inhibition / COS-7	(Cavalli; Babey et al. 1999)
OPRD_mouse	delta opioid receptor	TM3	137 LMSLDDECLAC A	IP production / COS-7	(Fanelli, Barbour et al. 1999)
OXYR_human	oxytocin	IC2			

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<b>PAFR_human</b>	platelet-activating factor (PAF)	C-terminal of IC3	231 <b>EVKRRALWWVCTVLAV</b> R	IP production / COS-7	(Parent, Le G uill et al. 1996)
<b>PAFR_human</b>	platelet-activating factor (PAF)	TMIII	100 <b>CLFFINTYCSV</b> A	arachidonate release, IP production, adenylyl cyclase inhibition / CHO	(Ishii, Izumi et al. 1997)
<b>PE23_human</b>	prostaglandin E <sub>3</sub> , EP3III, EP3IV	C-terminal tail	360 <b>FCQEEFWGN</b> <b>FCQMRKRRLRQQEEFWGN</b> ↑truncated	inhibition of adenylyl cyclase / CHO-K1	(Jin, Mao et al. 1997)
<b>PE23_mouse</b>	prostaglandin E <sub>3</sub> , EP3	carboxyl-terminal tail	336 <b>KILLRKFCQIRDHT</b> <b>MNNHL</b> ↑truncated	inhibition of adenylyl cyclase / CHO, stably expressed	(Hasegawa, Negishi et al. 1996)
<b>THRH_human</b>	thrombin	EC2 loop	259 CHDVLNNETLLEGKAYY DLKD KDF I	<sup>45</sup> Ca <sup>2+</sup> efflux, PI hydrolysis, reporter gene induction / COS-7	(Nanevicz, Wang et al. 1996)
<b>TSHR_human</b>	thyrotropin (TSHR) thyroid stimulating hormone	EC1 EC2	486 <b>YYNHAIIDWQTC</b> F,M 568 <b>YAKVSICLPMID</b> T	inositol phosphate-- diacylglycerol cascade / COS-7	(Parma, Van Sande et al. 1995)
<b>TSHR_human</b>	thyrotropin (TSHR) thyroid stimulating hormone	TMIII TMVII	509 <b>A S E L S Y V T I L T V</b> A 672 <b>Y P L N S C A N P F L</b> Y	adenylyl cyclase activation / COS-7	(Duprez, Parma et al. 1994)
<b>TSHR_human</b>	thyrotropin (TSHR) thyroid stimulating hormone	TMV	597 <b>V A F V I Y C C C H V</b> L	cAMP formation / COS-7 cells	(Esapa, Duprez et al. 1999)
<b>TSHR_human</b>	thyrotropin (TSHR) thyroid stimulating hormone	TMVII	677 <b>C A N P F L Y A I F T</b> V	cAMP formation / CHO cells	(Russo, Wong et al. 1999)
<b>TSHR_human</b>	thyrotropin (TSHR) thyroid stimulating hormone	IC3	613      621 <b>V R N P Q O Y N P G D K D I K A K</b> deletion	cAMP formation / COS-7	(Wonenow, Schoneberg et al. 1998)

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TSHR_human	thyrotropin (TSHR) thyroid stimulating hormone	IC3 / TMVI IC2	623      632 KDTKIAKRMMAVLIFIDPFICM V            I 136 LAMTLDQRHRAI A	cAMP activation / COS-7 cAMP formation / COS-7	(Paschke, Tonacchera et al. 1994) (Morin, Cotte et al. 1998)
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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS B GROUP I CALR_human	human calcitonin hCTR-1 hCTR-2	wild type (native) protein		adenylyl cyclase cAMP production / COS-1	(Cohen, Thaw et al. 1997)
CLASS B GROUP II PTRK_human	parathyroid hormone PTH / PTH-related peptide	junction of IC1 and TMII	223 TRNYIHMHLFL R,K	cAMP accumulation / COS-7	(Schipani, Jensen et al. 1997)
		junction of IC3 and TMVI	410 KLIKSTLVLMPC C,others		
CLASS B GROUP III GIPR_human	glucose-dependent insulinotropic peptide (GIP-R)	TMVI	340 VFAPVTEEQAR P	cAMP production / L293	(Tseng and Lin 1997)
GLR_rat	glucagon	junction of IC loop1 and TMII	178 TRNYIHGNLFA R	cAMP accumulation / COS-7	(Hjorth, Orskov et al. 1998)
		IC end of TMVI	352 RLARSTLILP A		
VIPR_human	vasoactive intestinal peptide 1 (VIP)	junction of IC loop 1 and TMII	178 RNYYIHMHLFFI R	cAMP production / COS-7 or CHO	(Gaudin, Maoret et al. 1998)
		junction of IC loop 3 and TMVI	343 LARSTLILP X= K,P		(Gaudin, Rouyer-Fessard et al. 1998)

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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS_C			TLSFVAQNKPSIANDRRCNCSEHIT		
CASR_human	calcium-sensing N-terminal EC	various substitutions, in multiple combinations	IP / tsA	(Jensen, Spalding et al. 2000)	

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File Name	Receptor	Mutation Site	Sequence	Assay / Cells	Reference
CLASS_D	pheromone	TM6	229 PLSAYQIVLGT P	heterologous yeast assay	(Olesnický, Brown et al. 1999)
RCB2					
C_cinerarius					
STE2_yeast	pheromone $\alpha$ -factor	TM6	258 QSLIVVPSIIFI LL	<i>lacZ</i> reporter gene	(Konopka, Margarit et al. 1996)
STE2_yeast	pheromone $\alpha$ -factor	double mutations TM5 and TM6	223 MSFVLYVKILAIR C C 247 251 DSFHILLIICOSLL CC CC	<i>lacZ</i> reporter gene / yeast	(Dube, DeCostanzo et al. 2000)
			double mutations		
			double mutations		
STE3_yeast	pheromone $\alpha$ -factor	IC3	194 DVRDLILACTNS Q	$\beta$ -galactosidase	(Boone, Davis et al. 1993)
STE2_yeast	pheromone $\alpha$ -factor	TM6	253 258 LIMSCQSLIVVPSIIFI L L P	$\beta$ -galactosidase	(Sommers, Martin et al. 2000)

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# A Point Mutation Enhances MC-4 Receptor Constitutive Activity

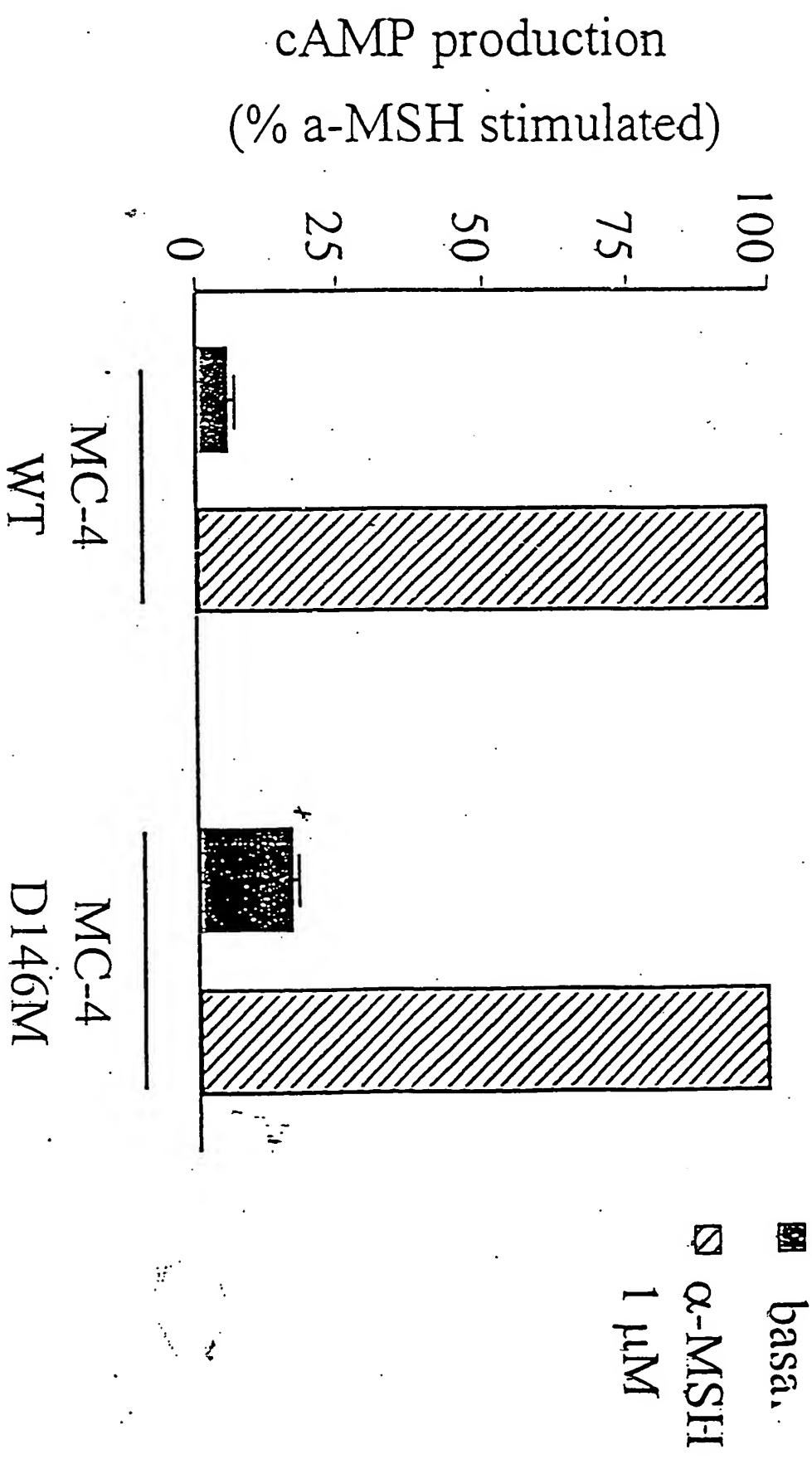


Figure 2

TOSONT™ SILENT™  
Light Emission Induced by the WT CCK-BR  
vs. a Constitutively Active Mutant

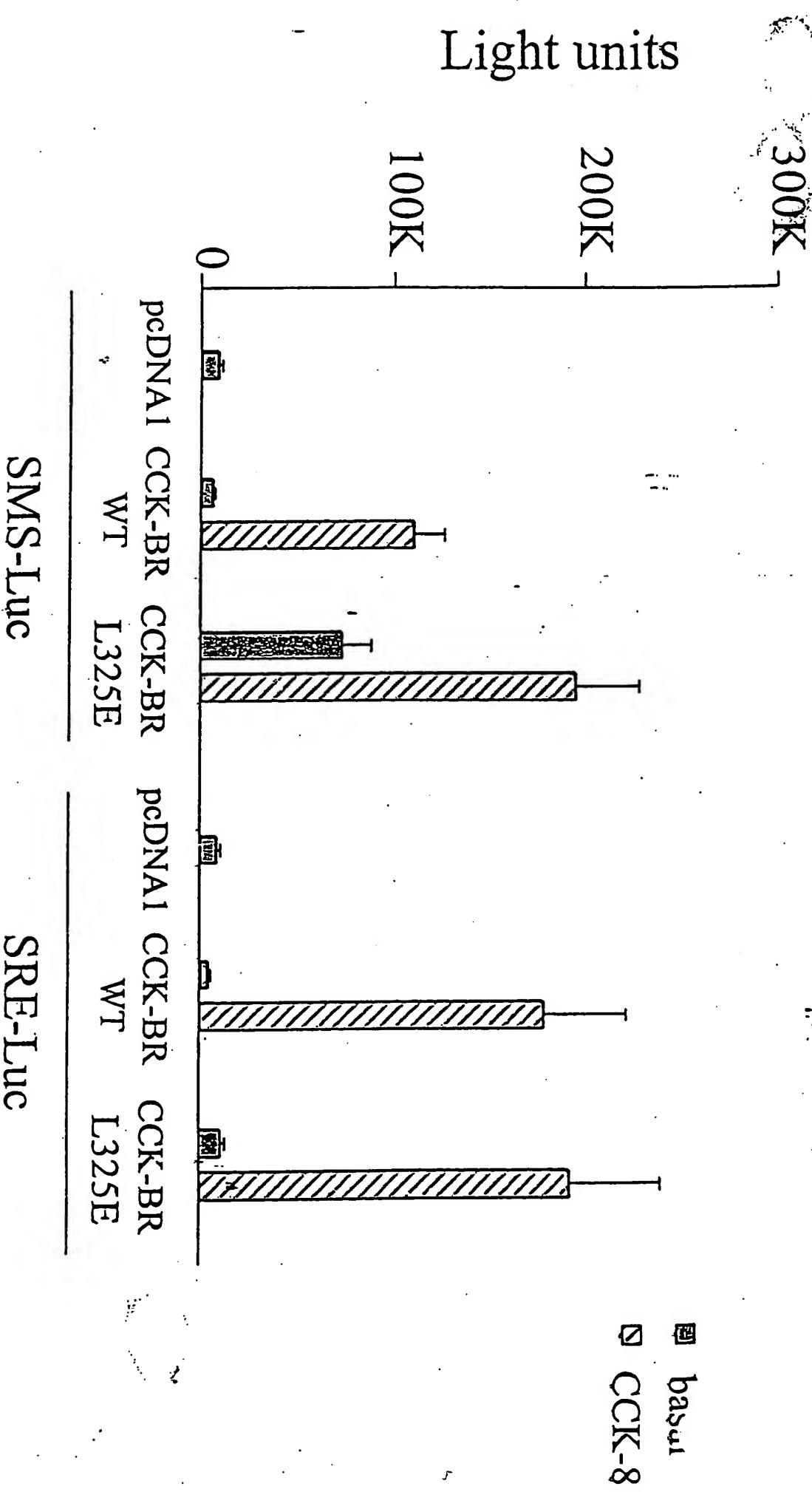


Figure 3

# A Point Mutation Confers Constitutive Activity to the Rat $\mu$ Opiod Receptor

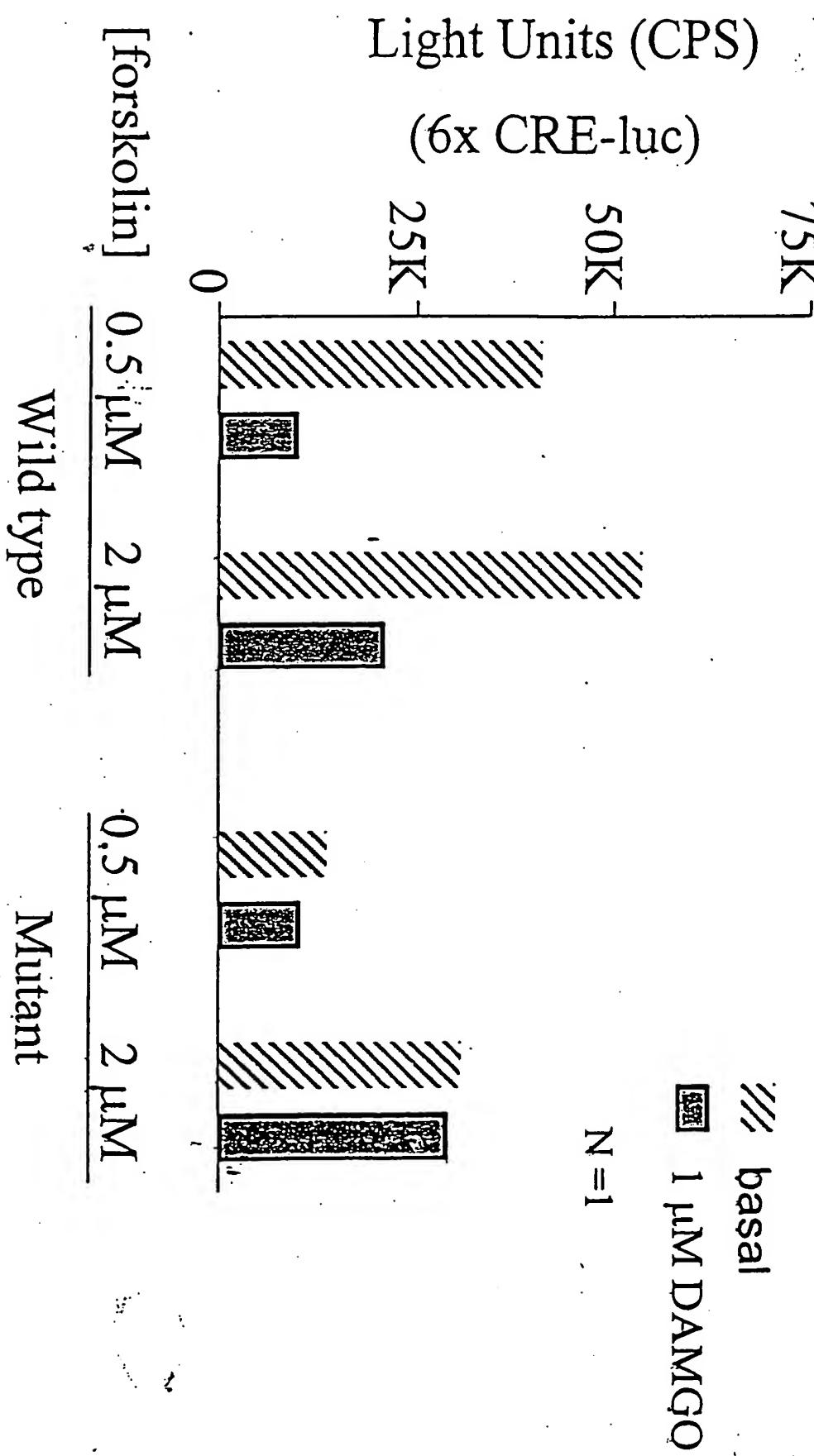


Figure 4

# Forskolin Stimulated HEK293 Cells Transfected With pcDNA1 and a CRE-luc Construct

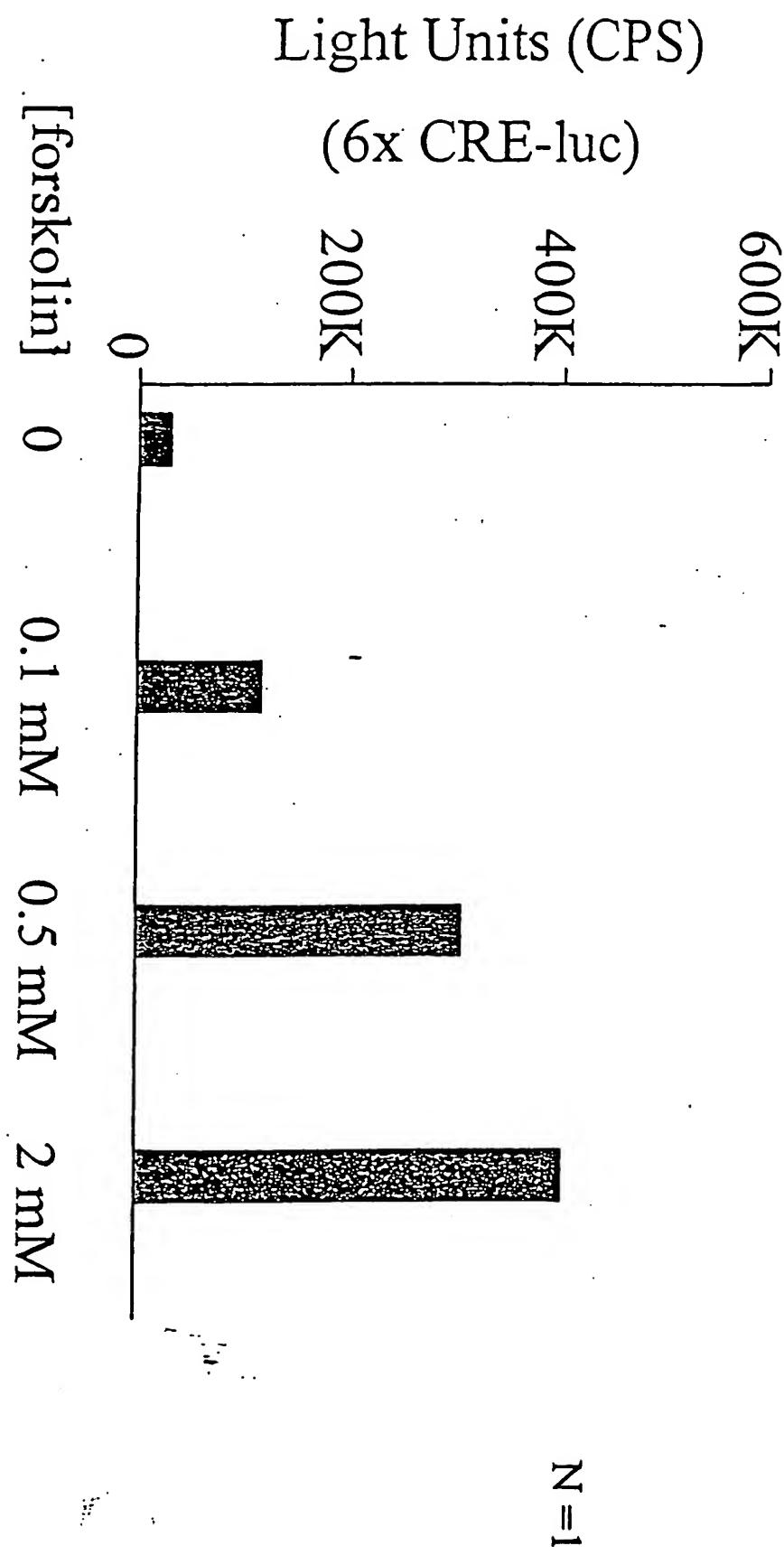


Figure 5

# The Rat $\mu$ Opioid Receptor Signals Through G $\alpha$ i

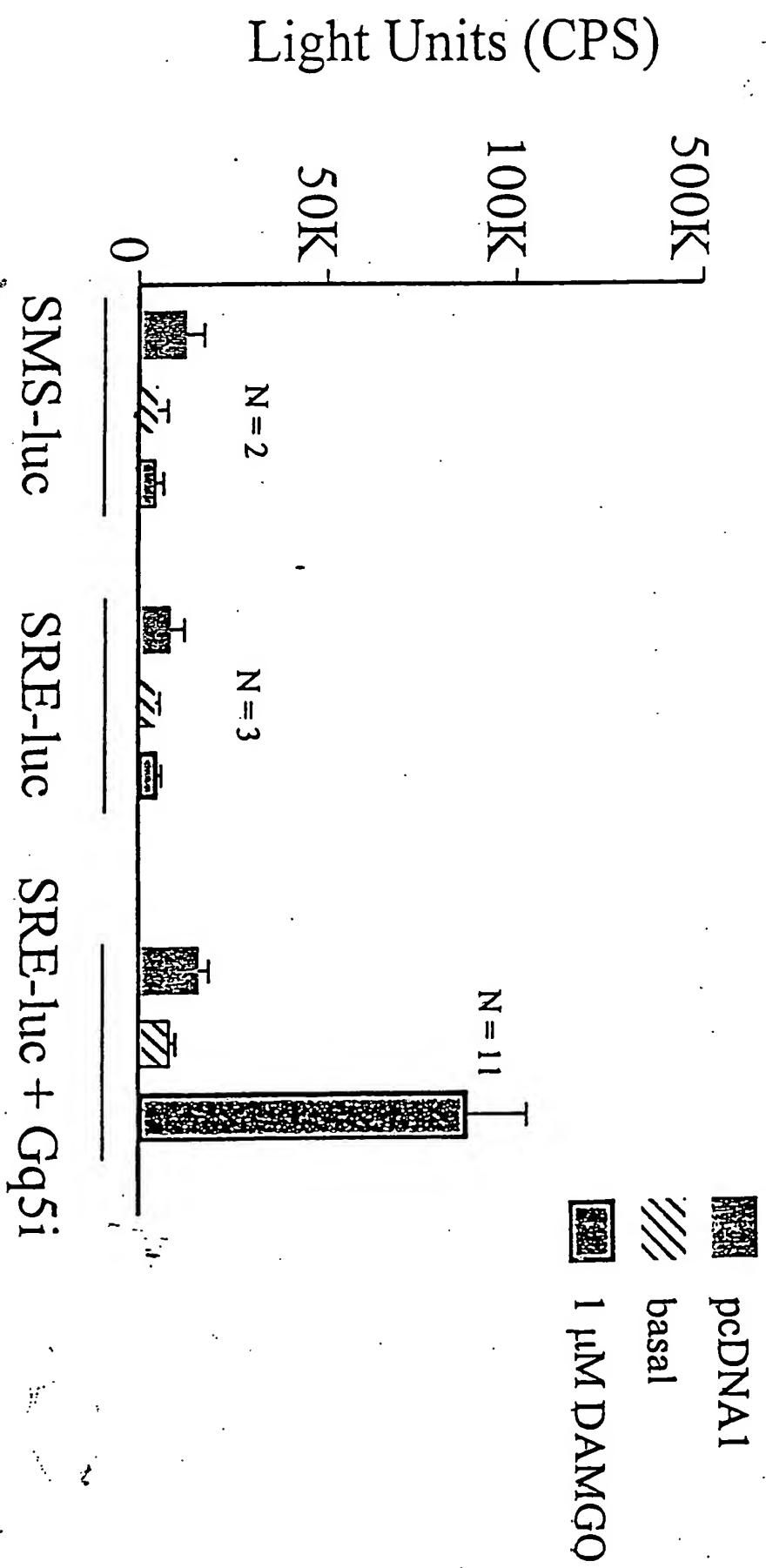


Figure 6

T O S 2 0 T " C H 0 6 E O G T

# A Point Mutation Confers Constitutive Activity to the Rat $\mu$ Opioid Receptor

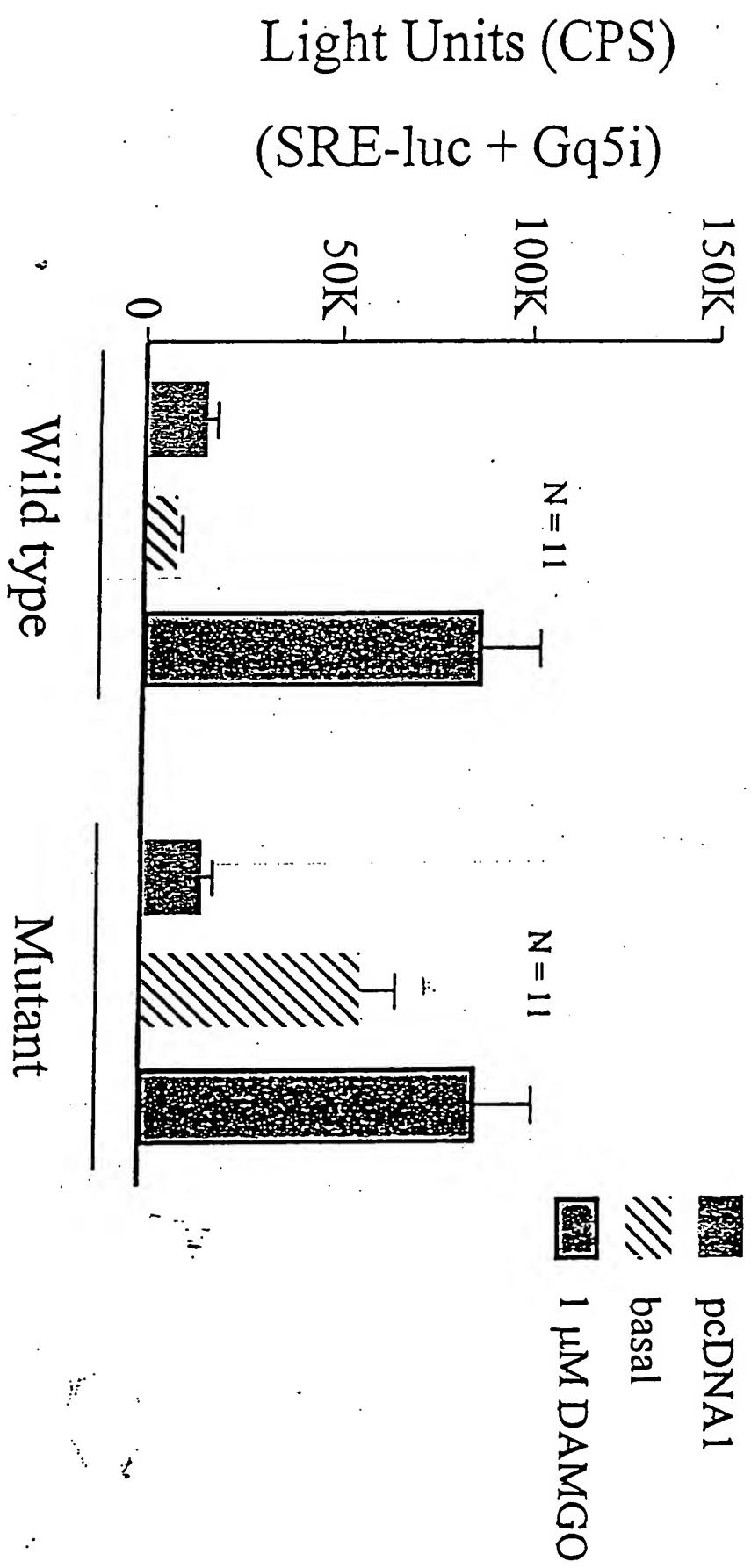


Figure 7

# Target Residues Within Class I GPCRs

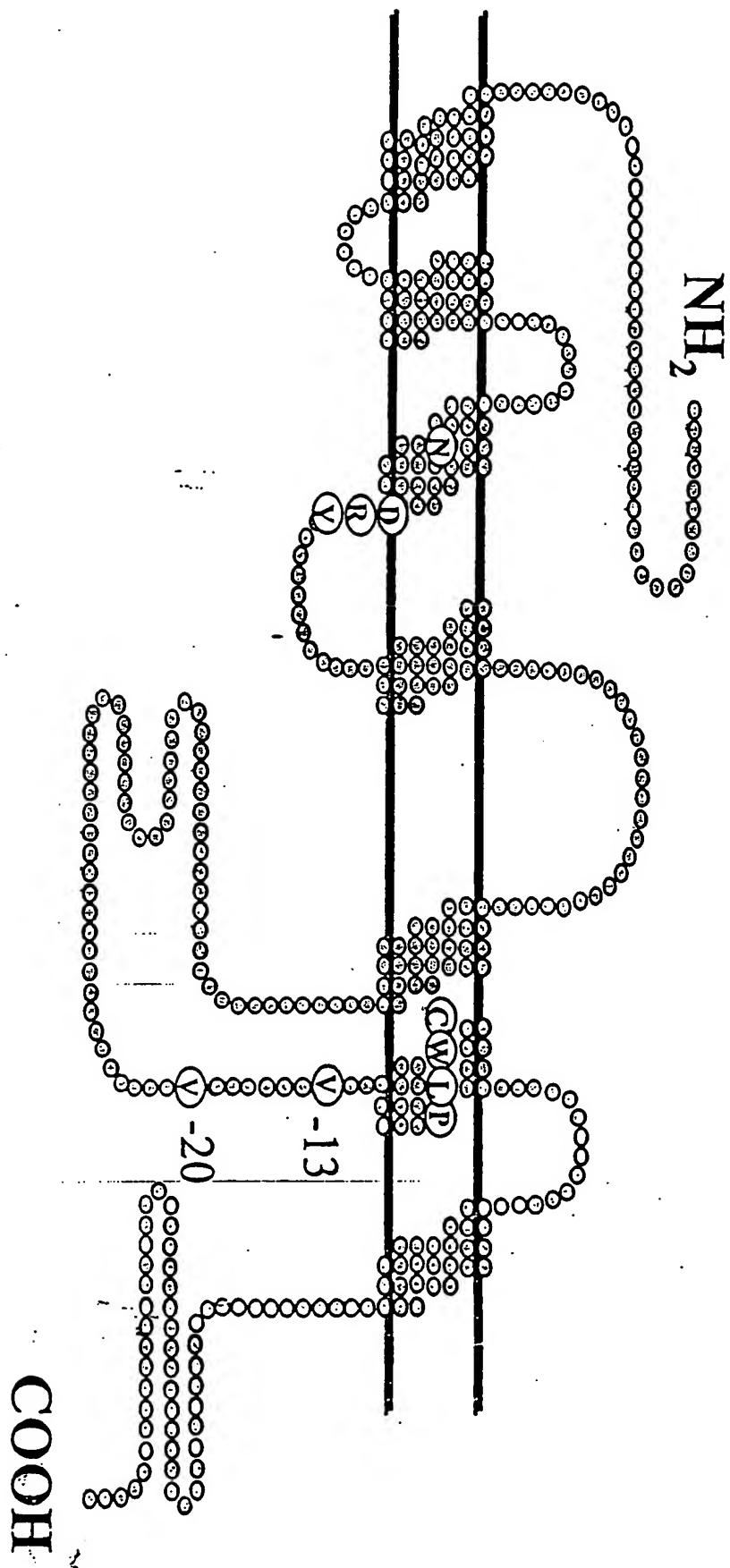


Figure 8

F O S S I L " C H A S E D "

# TMD III Asn (-14 from DRY) is a Target for Mutation Induced Constitutive Activity

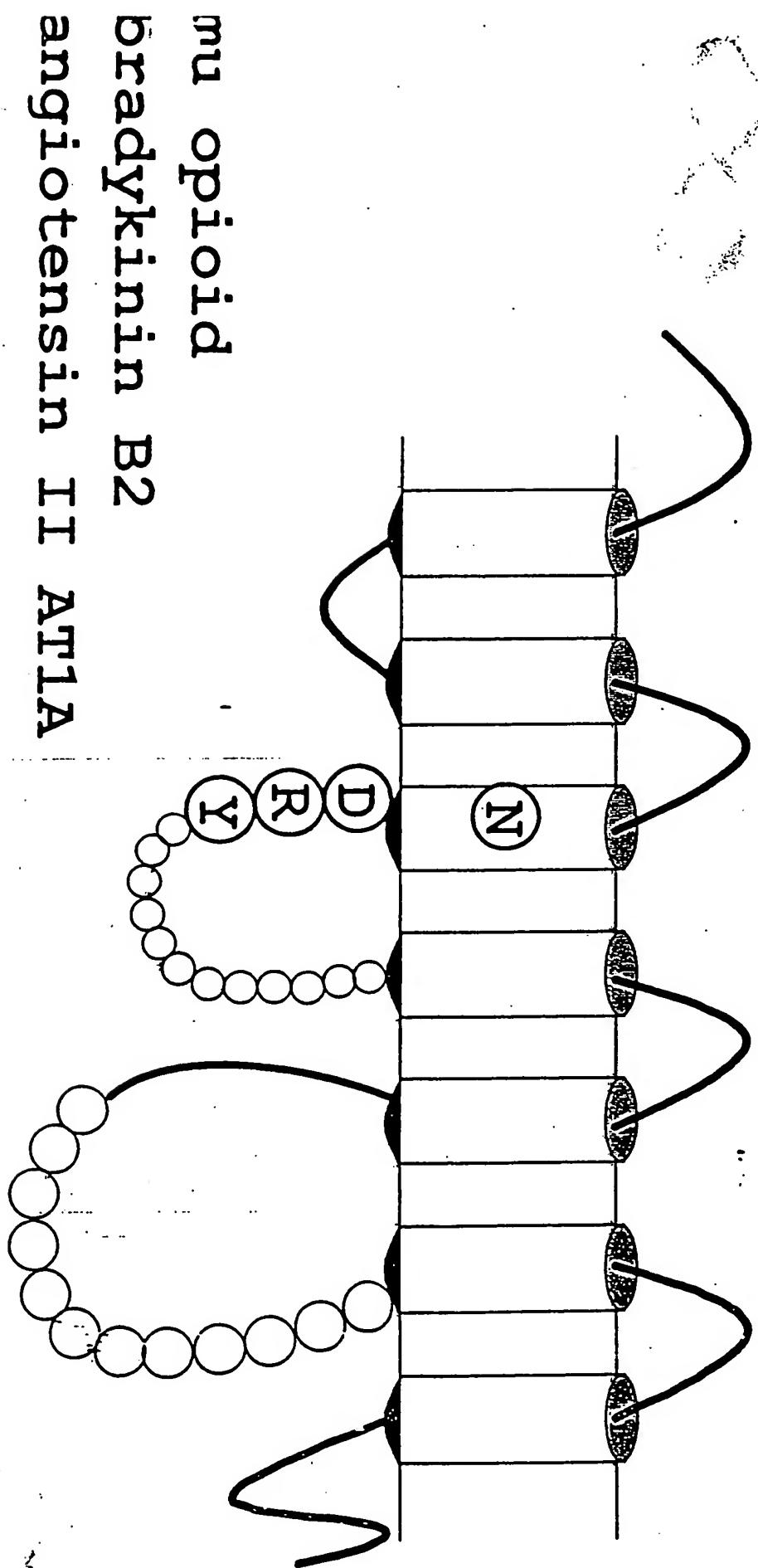


Figure 9

mu opioid  
bradykinin B2  
angiotensin II AT1A

# The 'DRY' Motif is a Target for Mutation Induced Constitutive Activity

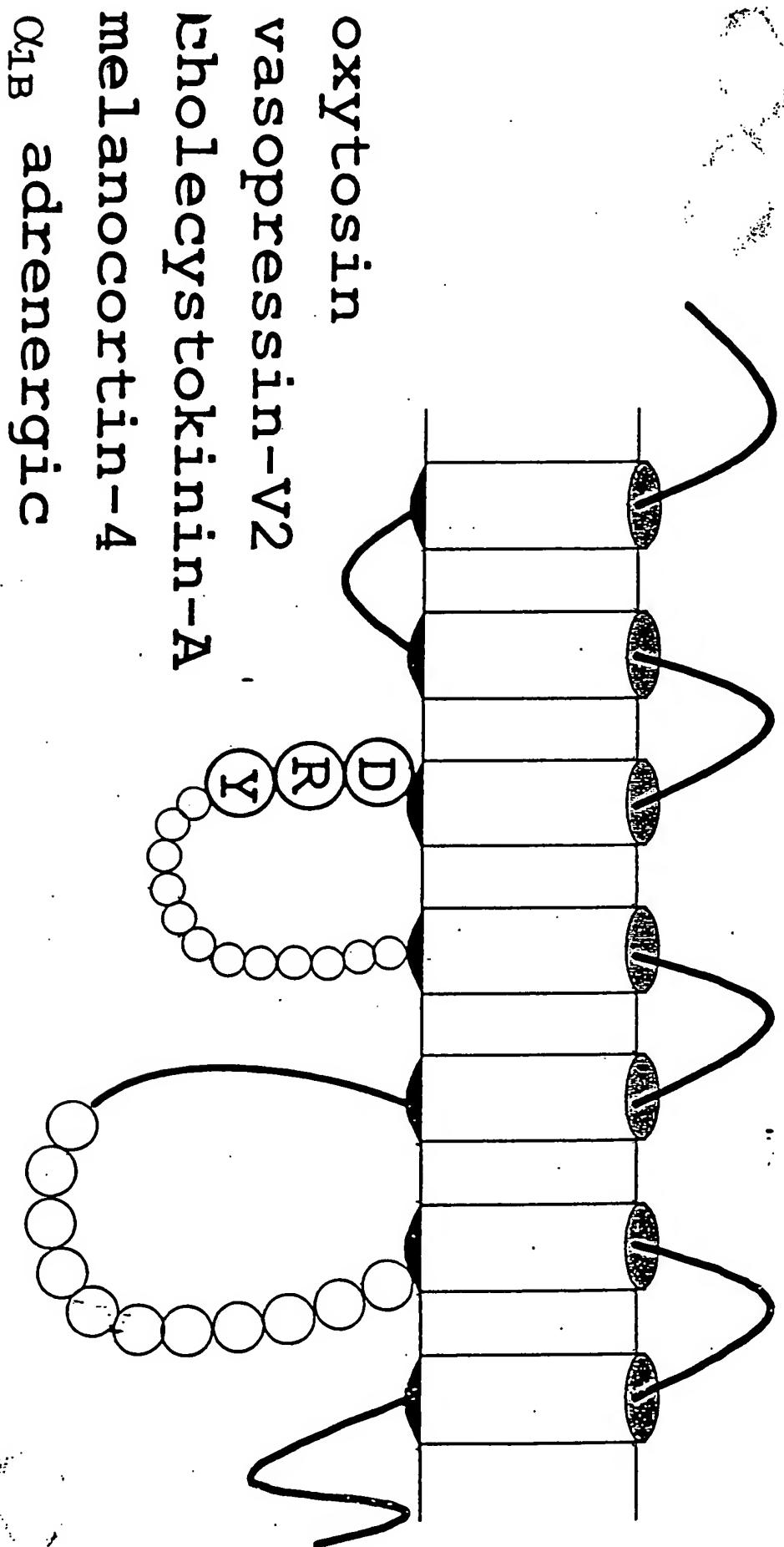


Figure 10

"R0520T" 5426E00T

## A Point Mutation Enhances MC-4 Receptor Constitutive Activity

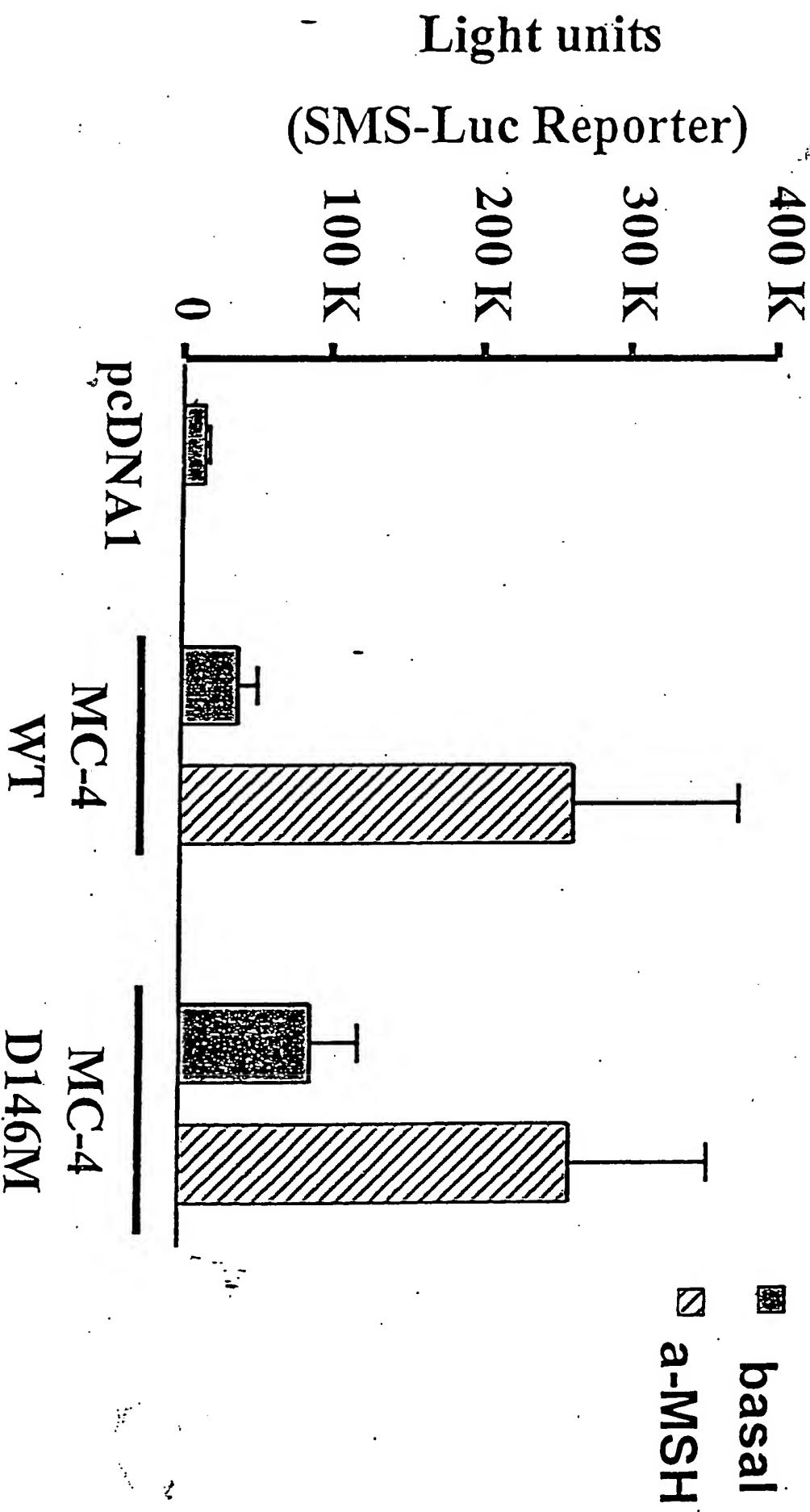


Figure 11

# The -13 Position is a Target for Mutation Induced Constitutive Activity

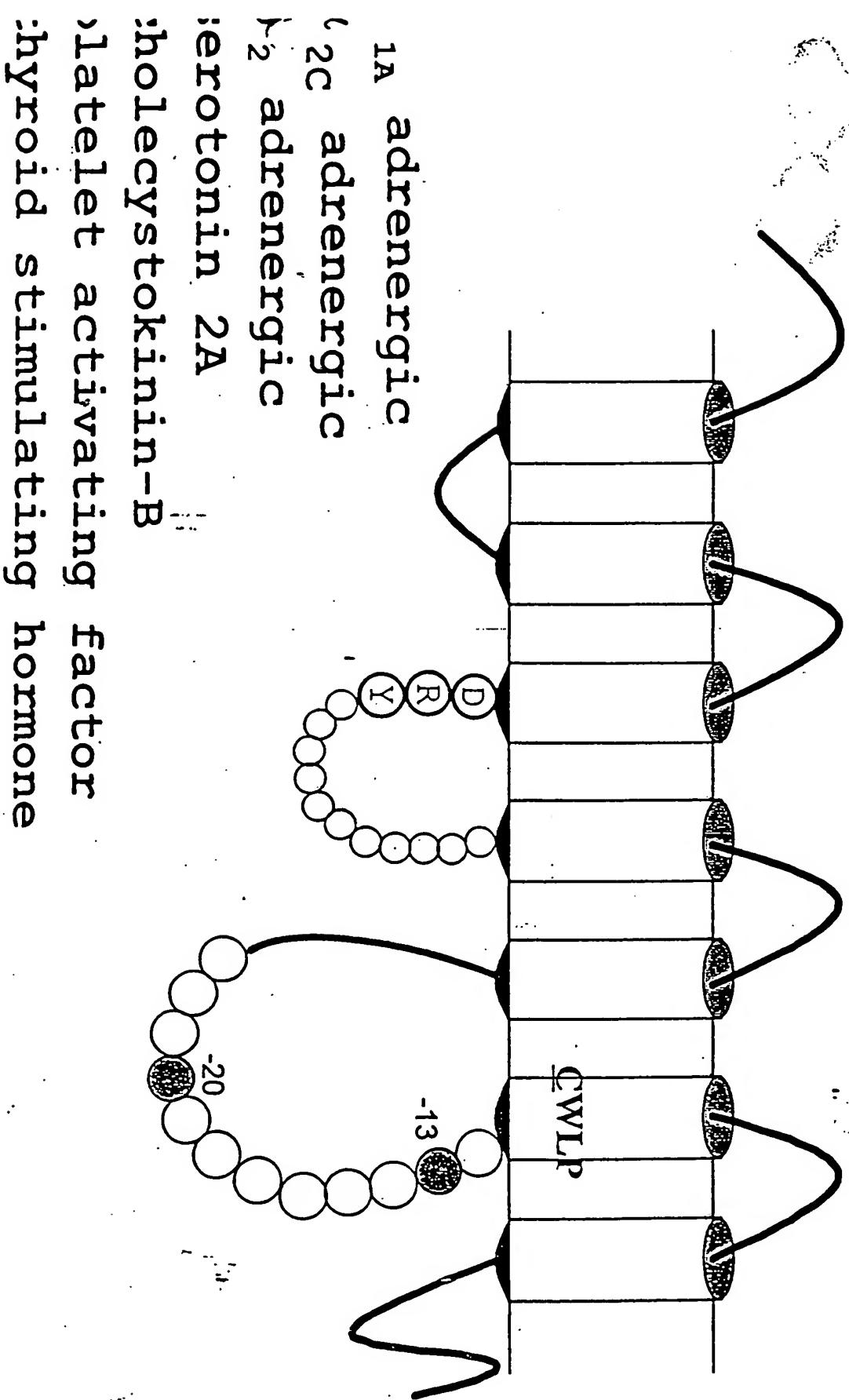


Figure 12

**Figure 13**



# Luciferase Activity (% maximum stimulation)

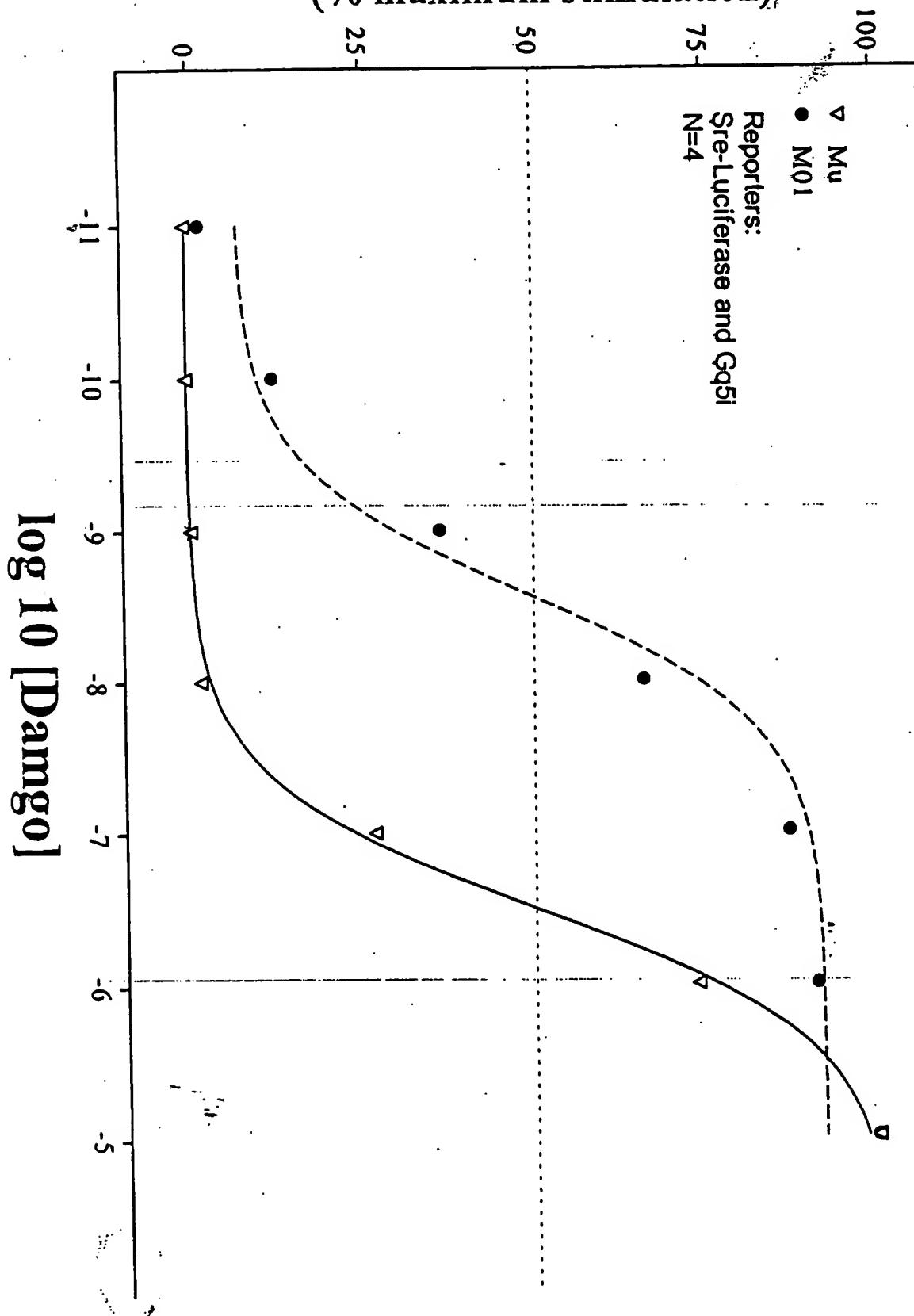


Figure 15

TO S E T " S H I G E O O T

# An Intracellular Point Mutation Results in Loss of Ligand-Induced Function

IP Production /  $^3\text{H}$  Inositol incorporation

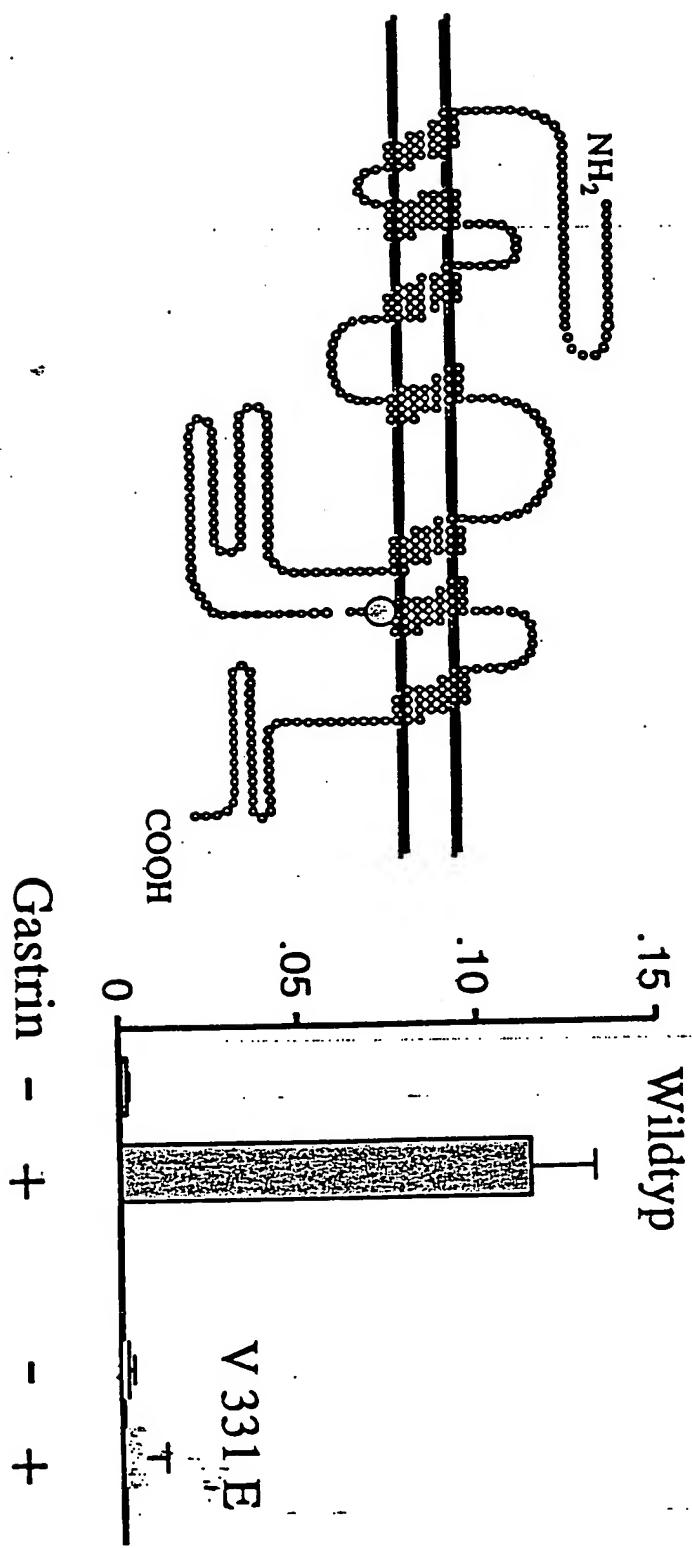


Figure 16

Figure 17

